

# Sphingosine 1-phosphate has a negative effect on RBC storage quality

Ariel Hay,<sup>1,\*</sup> Travis Nemkov,<sup>2,\*</sup> Fabia Gamboni,<sup>2</sup> Monika Dzieciatkowska,<sup>2</sup> Alicia Key,<sup>2</sup> Matthew Galbraith,<sup>3</sup> Kyle Bartsch,<sup>3</sup> Kaiqi Sun,<sup>4</sup> Yang Xia,<sup>5</sup> Mars Stone,<sup>6,7</sup> Michael P. Busch,<sup>6,7</sup> Philip J. Norris,<sup>6,7</sup> James C. Zimring,<sup>1</sup> and Angelo D'Alessandro<sup>2</sup>

<sup>1</sup>Department of Pathology, University of Virginia, Charlottesville, VA; <sup>2</sup>Department of Biochemistry and Molecular Genetics, and <sup>3</sup>Linda Crnic Institute for Down Syndrome, Anschutz Medical Campus, University of Colorado, Aurora, CO; <sup>4</sup>Metis Therapeutics, Shanghai, China; <sup>5</sup>University of Changsha, Changsha, China; <sup>6</sup>Vitalant Research Institute, San Francisco, CA; and <sup>7</sup>Department of Laboratory Medicine, University of California, San Francisco, CA

## Key Points

- S1P declines during RBC storage. S1P supplementation to human or murine RBCs boosts energy metabolism at the expense of antioxidant pathways.
- S1P worsens and Sphk1 KO, or hypoxia, promote stored RBC posttransfusion recoveries.

Blood storage promotes the rapid depletion of red blood cell (RBC) high-energy adenosine triphosphate (ATP) and 2,3-diphosphoglycerate (DPG), which are critical regulators of erythrocyte physiology and function, as well as oxygen kinetics and posttransfusion survival. Sphingosine-1-phosphate (S1P) promotes fluxes through glycolysis. We hypothesized that S1P supplementation to stored RBC units would improve energy metabolism and posttransfusion recovery. We quantified S1P in 1929 samples (n = 643, storage days 10, 23, and 42) from the REDS RBC Omics study. We then supplemented human and murine RBCs from good storer (C57BL6/J) and poor storer strains (FVB) with S1P (1, 5, and 10  $\mu$ M) before measurements of metabolism and posttransfusion recovery. Similar experiments were repeated for mice with genetic ablation of the S1P biosynthetic pathway (sphingosine kinase 1 [Sphk1] knockout [KO]). Sample analyses included metabolomics at steady state, tracing experiments with 1,2,3-<sup>13</sup>C<sub>3</sub>-glucose, proteomics, and analysis of end-of-storage posttransfusion recovery, under normoxic and hypoxic storage conditions. Storage promoted decreases in S1P levels, which were the highest in units donated by female or older donors. Supplementation of S1P to human and murine RBCs boosted the steady-state levels of glycolytic metabolites and glycolytic fluxes, ie the generation of ATP and DPG, at the expense of the pentose phosphate pathway. Lower posttransfusion recovery was observed upon S1P supplementation. All these phenomena were reversed in Sphk1 KO mice or with hypoxic storage. S1P is a positive regulator of energy metabolism and a negative regulator of antioxidant metabolism in stored RBCs, resulting in lower posttransfusion recoveries in murine models.

## Introduction

Red blood cell (RBC) transfusion is a life-saving intervention for millions of recipients worldwide every year. However, the quality of packed RBC products declines during storage in the blood bank, which promotes a series of morphological<sup>1,2</sup> and biochemical changes<sup>3</sup> that ultimately affect erythrocyte physiology and posttransfusion performances. The quality of stored RBCs, as per the US Food and Drug Administration (FDA) and the European Council guidelines, is defined by hemolytic propensity and the capacity of transfused RBCs to circulate at 24 hours after transfusion (henceforth, posttransfusion

Submitted 12 September 2022; accepted 12 November 2022; prepublished online on *Blood Advances* First Edition 5 December 2022; final version published online 12 April 2023. <https://doi.org/10.1182/bloodadvances.2022008936>.

\*A.H. and T.N. contributed equally to this study and are joint first authors.

Data are available on request from the corresponding author, Angelo D'Alessandro ([angelo.dalessandro@cuanschutz.edu](mailto:angelo.dalessandro@cuanschutz.edu)).

The full-text version of this article contains a data supplement.

© 2023 by The American Society of Hematology. Licensed under [Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International \(CC BY-NC-ND 4.0\)](https://creativecommons.org/licenses/by-nc-nd/4.0/), permitting only noncommercial, nonderivative use with attribution. All other rights reserved.

recovery [PTR]); these 2 parameters increase and decrease, respectively, as a function of storage duration.<sup>3</sup>

It has been argued that oxidative stress is the main driver of the storage lesion.<sup>4</sup> RBCs are well equipped to counteract oxidant stress through different systems, among which the pentose phosphate pathway (PPP) represents a critical lynchpin. The PPP is the main pathway generating the key reducing equivalent nicotinamide adenine dinucleotide phosphate, NADPH, which participates in the scavenging of reactive oxygen species and recycling of oxidized soluble small molecules (for example, the glutathione system) and enzyme-catalyzed antioxidant batteries.<sup>5</sup> Glucose-6-phosphate dehydrogenase (G6PD) is the rate-limiting enzyme of the PPP, and it is highly polymorphic in humans.<sup>6</sup> Of note, antioxidant capacity,<sup>7</sup> storage quality,<sup>8</sup> hemolytic propensity,<sup>9</sup> PTRs,<sup>10</sup> and posttransfusion hemoglobin increments<sup>11</sup> are poorer in packed RBCs donated by volunteers suffering from G6PD deficiency, a condition that affects ~500 million individuals to a variable extent, owing to a plethora of genetic mutations, all resulting in a hypomorphic or unstable G6PD enzyme.<sup>6</sup> Because G6PD is a chromosome X-linked gene, whose activity declines with age<sup>12</sup> and affects lifespan in mammals,<sup>13</sup> it is interesting to note that donor sex and age<sup>8,14,15</sup> affect RBC antioxidant capacity and posttransfusion RBC performances beyond the chronological age of the unit, that is, the days elapsed since the time of donation.<sup>16,17</sup>

Others have noted that rapid depletion of high-energy phosphate compounds (adenosine triphosphate [ATP] and 2,3-diphosphoglycerate [DPG]) is a hallmark of the RBC metabolic storage lesion.<sup>18</sup> ATP consumption is accompanied by dysregulated calcium ion pumps,<sup>19</sup> resulting in intracellular calcium accumulation and eryptosis.<sup>20</sup> Reductions in ATP deprive erythrocytes of a rate-limiting substrate for phosphorylation of structural membrane proteins, which is critical to morphological homeostasis of the stored RBC.<sup>21</sup> By fueling ATP-dependent flippases, ATP depletion in the stored RBC results in phosphatidylserine exposure to the outer membrane leaflet and untimely removal upon transfusion.<sup>3,22,23</sup> DPG exhaustion by storage week 2 to 3 is mechanistically linked to altered oxygen kinetics.<sup>22</sup> Indeed, depletion of DPG levels promotes increases in oxygen saturation (SO<sub>2</sub>) by shifting the oxygen dissociation curve to the left,<sup>24</sup> which in turn promotes the concomitant storage-dependent formation of reactive oxygen species<sup>2</sup> that are triggered by Haber-Weiss chemistry in the presence of elevated O<sub>2</sub> levels. Indeed, heterogeneity in baseline SO<sub>2</sub> levels after processing is associated with heterogeneity in storage quality,<sup>25</sup> to the extent that SO<sub>2</sub> manipulation by hypoxic storage improves energy metabolism, prevents the oxidative storage lesion,<sup>26,27</sup> and boosts PTRs in randomized clinical trials in humans,<sup>28</sup> as well as transfusion efficacy in rodent models of trauma and shock.<sup>29</sup> Among the mechanisms identified as key drivers of the benefits of hypoxic storage is the alkalization of intracellular pH,<sup>30</sup> and the mitigation of cysteine oxidation<sup>27</sup> and asparagine deamidation<sup>26</sup> of key glycolytic enzymes. These factors, combined with deoxyhemoglobin competitive binding to the N-terminus of band 3 that displaces otherwise bound/inhibited glycolytic enzymes,<sup>31-33</sup> all contribute to boosting glycolysis and ATP generation capacity in hypoxically stored RBCs, which ultimately preserves oxygen kinetics of these products.<sup>34</sup> Of note, even though DPG and ATP synthesis is slowly restored upon transfusion in vivo,<sup>35</sup> the kinetics may be insufficient to meet the

oxygen requirements in the hypoxic, massively transfused recipient (for example, patients with trauma).

In previous mechanistic studies on high-altitude hypoxia and sickle cell disease,<sup>36,37</sup> we showed that sphingosine-1-phosphate (S1P) supplementation promotes fluxes through glycolysis by mediating hemoglobin binding to the N-terminus domain of band 3,<sup>37</sup> thereby displacing glycolytic enzymes, otherwise inhibited by competitive binding to the very same domain of band 3.<sup>32,33</sup> By promoting glucose oxidation through glycolysis and the Rapoport-Luebering shunt, S1P supplementation promotes the synthesis of ATP and DPG. Exposure to hypoxia (in vivo or in vitro) promotes S1P synthesis by RBC sphingosine kinase 1 (Sphk1), which is critical to metabolic adaptations to high-altitude or pathological hypoxia (for example, chronic kidney disease).<sup>37,38</sup> However, previous studies have shown that storage is accompanied by declines in S1P levels (down to 19% of fresh values in day-30 units<sup>39</sup>), suggesting that exogenous supplementation of S1P could be leveraged as a metabolic intervention in stored RBC units. Other studies, however, have either shown no storage-dependent change<sup>40</sup> or additive solution-dependent increases/decreases in S1P levels.<sup>41</sup> In light of this, first of all, we leveraged the largest sample set amenable for S1P assessment from the REDS RBC Omics study,<sup>42</sup> a longitudinal study on ~2000 samples from 643 blood donors. After confirming that storage causes a decline in S1P levels in RBC units, especially those donated by donors characterized by extreme hemolytic propensity from the REDS RBC Omics cohort,<sup>42</sup> we hypothesized that supplementation of S1P to human and murine packed RBCs would improve storage quality, whereas genetic ablation of Sphk1 would impair S1P biosynthesis and, consequently, exacerbate the storage lesion by negatively affecting glycolysis (and thus ATP and DPG synthesis).

## Methods

### REDS RBC Omics study participants and samples

RBC Omics was conducted under regulations applicable to all human subject research supported by federal agencies as well as the requirements for blood product manipulation specified and approved by the FDA. The data coordinating center (RTI International) of REDS was responsible for the overall compliance of human participant regulatory protocols including institutional review board approval from each participating blood center, from the REDS Central Laboratory (Vitalant Research Institute), and from the data coordinating center, as previously detailed.<sup>8,41</sup> Donors were enrolled at the 4 participating REDS US blood centers. Overall, 13 403 individuals aged ≥18 years provided informed consent to participate in the study. Hemolysis parameters (spontaneous, oxidative, or osmotic) were evaluated on stored RBCs from these donors after ~39 to 42 days of storage. Extreme hemolyzers (5th and 95th percentile) from the donors tested for end-of-storage oxidative hemolysis were asked to donate a second unit of blood. These units were sterilely sampled for metabolomics analysis (n = 643, storage days 10, 23, and 42). Blood collection, sample processing, and other aspects of the screening and recall phases of the RBC Omics Study have been extensively described.<sup>9,14</sup>

### Human RBC incubation with D7-S1P

To validate S1P uptake by human erythrocytes,<sup>43,44</sup> RBCs (n = 8; 4 males and 4 females) were incubated at 37°C for 0, 3, 6, and

24 hours in AS-3 additive solution supplemented with the stable isotope-labeled D7-S1P (product #860659, Avanti Lipids) at 0, 1, 3, 6, or 12  $\mu\text{M}$ .

### RBC storage with S1P

Whole blood units were donated by 10 healthy donor volunteers in CP2D (Haemonetics, Boston, MA). Two pools of 5 units were made, upon leukofiltration, plasma removal, and suspension in AS-3 additive solution, with or without supplementation of S1P (product #MFCD00270077; CAS #26993-30-6; Sigma Aldrich, St. Louis, MO) at 1, 5, or 10  $\mu\text{M}$ .

### Mouse blood collection, storage under hypoxic and normoxic conditions, and PTR

All murine experimental protocols were approved by the University of Virginia IACUC on 22 April 2019 (protocol n: 4269). Mouse strains C57BL6/J, FVB and Sphk1 knockout (KO) have been previously described.<sup>45,46</sup> Murine RBC storage (for 3 and 6 days for FVB mice, and 12 days for C57BL6/J mice), transfusion, and PTR determinations were carried out as previously described, with minor modifications.<sup>46</sup> Whole blood was drawn by cardiac puncture under sterile conditions into CPDA-1, centrifuged, and the hematocrit was adjusted to 75% by removing supernatant. In a subset of experiments, units were supplemented with 5  $\mu\text{M}$  S1P. For hypoxic storage, murine RBCs were bubbled with nitrogen gas in a glove box until  $\text{SO}_2 < 50\%$  before storage, whereas normoxic counterparts reached  $\text{SO}_2 > 95\%$  by the end of the storage period. Sealed "units" were preserved at 37°C for 1 hour and stored at 4°C for 7 and 12 days. RBCs from C57BL6/J mice were used as the test population and subjected to different storage conditions. UBC-GFP mice (stock #004353) were used as recipients to allow visualization of the test cells in the nonfluorescent gate. To control for differences in transfusion and phlebotomy, RBCs from ROSA26-LCB-mCHERRY mice (mCHERRY) were used as a tracer RBC population (never stored) and were added to stored RBCs immediately before transfusion, as recently described.<sup>47</sup> PTR was calculated by dividing the posttransfusion ratio (test:tracer) by the pretransfusion ratio (test:tracer).<sup>48</sup> A single PTR value  $> 100\%$  was calculated across all replicates in all experiments, which was explained by the tracer population being damaged and/or outperformed by the test population. Because PTR  $> 100\%$  is not biologically meaningful, this value was set to an absolute value of 100%, as standard practice in the field. At the time of transfusion, blood samples were frozen in liquid nitrogen and stored at  $-80^\circ\text{C}$  until subsequent analysis.

### Glucose tracing experiments

RBCs (100  $\mu\text{L}$ ) from all the mouse strains investigated in this study were incubated at 37°C for 1 hour in the presence of 1,2,3- $^{13}\text{C}_3$ -glucose (5 mM, Cambridge Isotopes, product #CLM-4673) and stored for 7 and 12 days before determination of lactate isotopologues +2/+3 (as markers of PPP-to-glycolysis fluxes), as described.<sup>27</sup>

### Ultra-high-performance liquid chromatography (UHPLC)-mass spectrometry metabolomics

Frozen RBC aliquots (50  $\mu\text{L}$ ) were extracted 1:10 in ice cold extraction solution (methanol:acetonitrile:water, 5:3:2 v/v/v).<sup>26</sup>

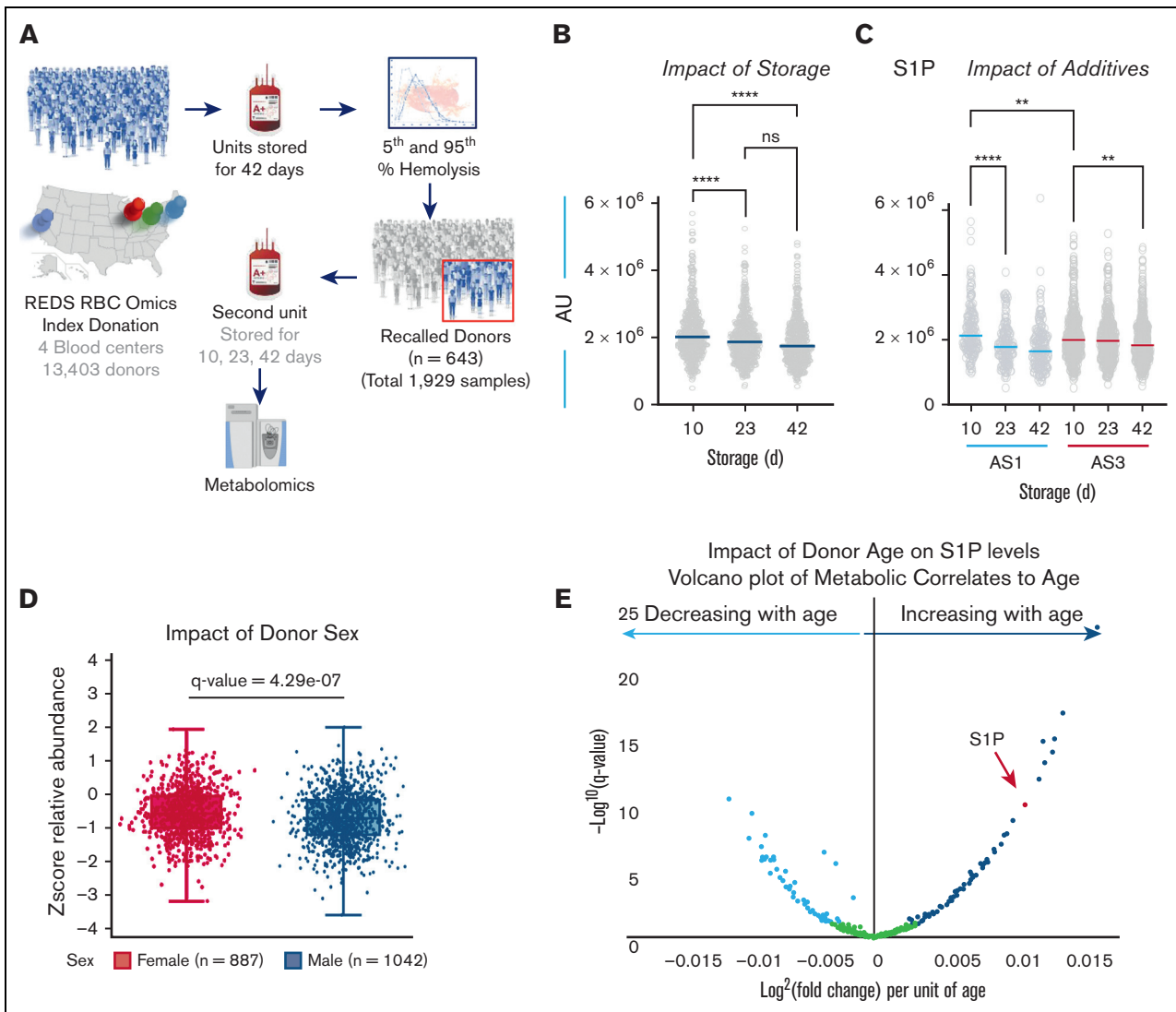
Samples were vortexed and insoluble material pelleted, as previously described.<sup>49</sup> Analyses were performed using a Vanquish UHPLC coupled online to a Q Exactive mass spectrometer (Thermo Fisher, Bremen, Germany). Samples were analyzed using a 1<sup>50</sup> and 5 minute gradient-based method,<sup>49</sup> as previously described.<sup>51,52</sup> S1P measurements were performed via the same platform described earlier, as validated in prior technical notes<sup>52</sup> and RBC-centric studies on S1P,<sup>37,38</sup> with the auxilium of stable isotope-labeled internal standards (D7-S1P, product #860659, Avanti Lipids). Data analysis was performed through the auxilium of the software MAVEN.<sup>53</sup> Graphs and statistical analyses (either two-way analysis of variance [ANOVA] or repeated-measures ANOVA) were prepared with GraphPad Prism 8.0 (GraphPad Software, Inc, La Jolla, CA), GENE E (Broad Institute, Cambridge, MA), and MetaboAnalyst 5.0.<sup>54</sup> All raw data are available from the corresponding author upon reasonable request.

## Results

### RBC S1P declines with storage and is higher in packed RBCs from female and older donors

As part of the REDS-III RBC Omics study, we analyzed packed RBC samples from 643 donors, at 3 storage time points (ie, days 10, 23, and 42; Figure 1A). These donors were selected among an original cohort of 13 403 healthy volunteers enrolled at 4 different blood centers across the United States and tested for RBC hemolytic propensity (spontaneous, following oxidant, or osmotic insults). Subjects whose RBCs tested in the 5th and 95th percentile for hemolytic propensity were contacted and invited to donate a second unit of blood, for a total of 1929 samples tested in this study. Measurements of S1P from packed RBC units in this study showed a progressive depletion of this metabolite as a function of storage duration (Figure 1B). Such measurements were performed via UHPLC-mass spectrometry, based on accurate intact mass and retention times validated against unlabeled or deuterium-labeled commercial standards (supplemental Figure 1A-B), which also confirmed RBC capacity to take up exogenous S1P in a dose-response manner (supplemental Figure 1C).

In the REDS-III RBC Omics cohort, S1P was nonnormally distributed across all blood donors at the end of storage (supplemental Figure 2A). Identification of donors in the bottom or top 5% based on S1P levels (supplemental Figure 2B) resulted in the identification of 2 separate groups across storage within the REDS-III RBC Omics cohort. Heat map representation of the top 50 metabolites based on donor S1P levels (supplemental Figure 2C) indicated a positive association between elevated S1P and glycolytic metabolites (fructose bisphosphate, bisphosphoglycerate, phosphoglycerate, phosphoenolpyruvate; supplemental Figure 3), markers of oxidant stress (glutathione disulfide, methionine sulfoxide), and several acylcarnitines, and a negative association with PPP metabolites (6-phosphogluconate, pentose phosphate). Notably, factors like additive solution, donor sex, and age were all associated with an effect on RBC S1P levels, independent of storage duration (Figure 1C-E). Specifically, RBCs stored in additive solution 1, from female and older donors were found to have higher levels of S1P (Figure 1C-E).

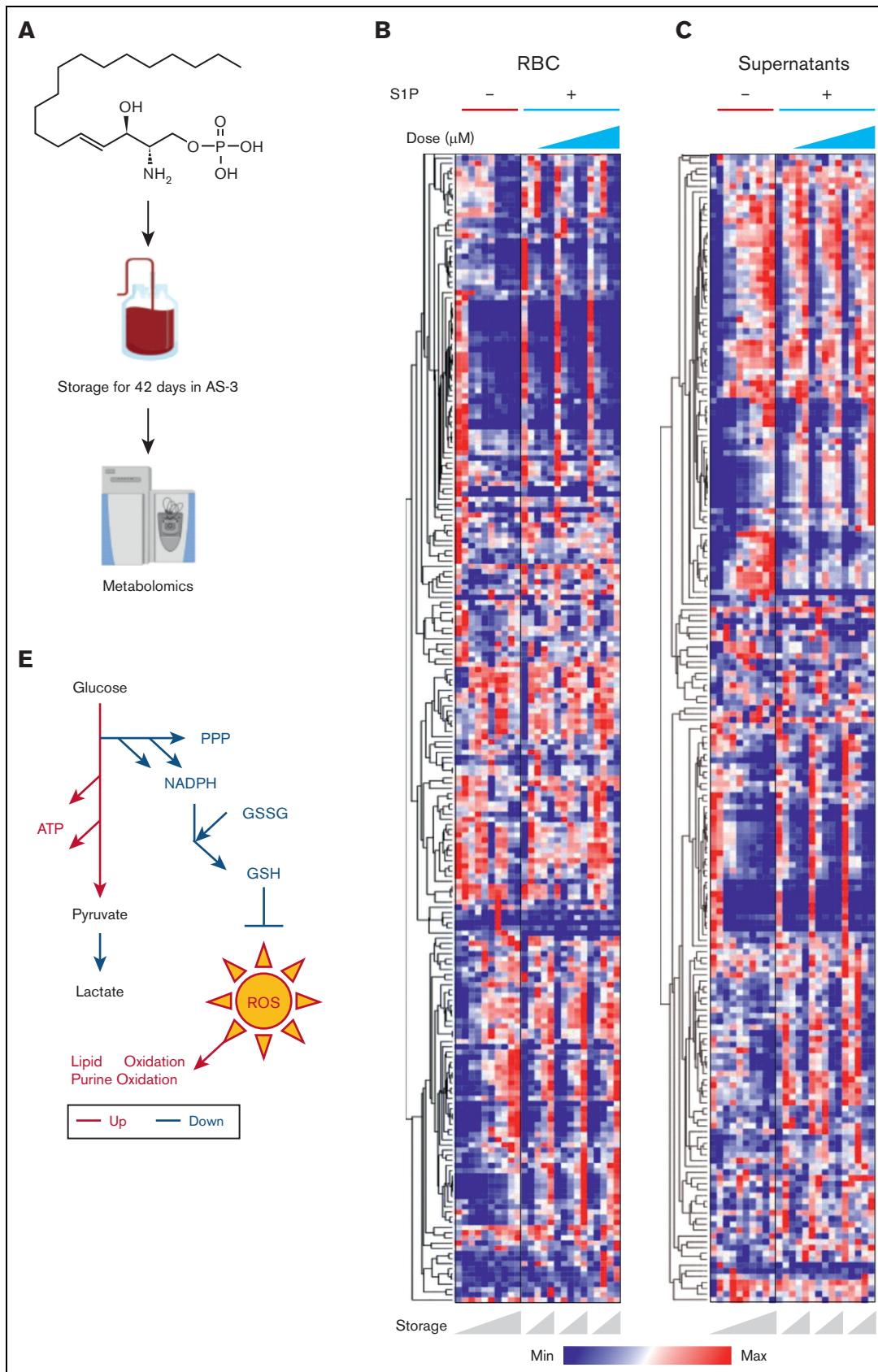


**Figure 1. RBC S1P decreases during storage and is higher in packed RBCs from female and older donors.** (A) An overview of the REDS-IV RBC Omics program is provided in panel A. (B-C) As part of the recalled donor arm of the study, 1929 units from 643 extreme hemolyzing donors were tested for RBC S1P levels (AUs, which were found to decrease during storage (B), and were lower early on during storage in AS-3 packed RBC units (AU) (C). (D-E) Higher levels of S1P were observed in blood donated by female donors (box and whisker plot; median  $\pm$  ranges[D]), older donors (volcano plot of metabolic correlates to age across all blood donors tested in this study; x-axis indicates log<sub>2</sub>-fold changes per unit of age for each metabolite and the y-axis indicates the negative log<sub>10</sub> of q-values for such correlations [E]), independent storage duration and additive. Asterisks indicate significance (ANOVA with multiple column comparisons; \*\**P* < .01, \*\*\**P* < .001, \*\*\*\**P* < .0001). AU, arbitrary unit.

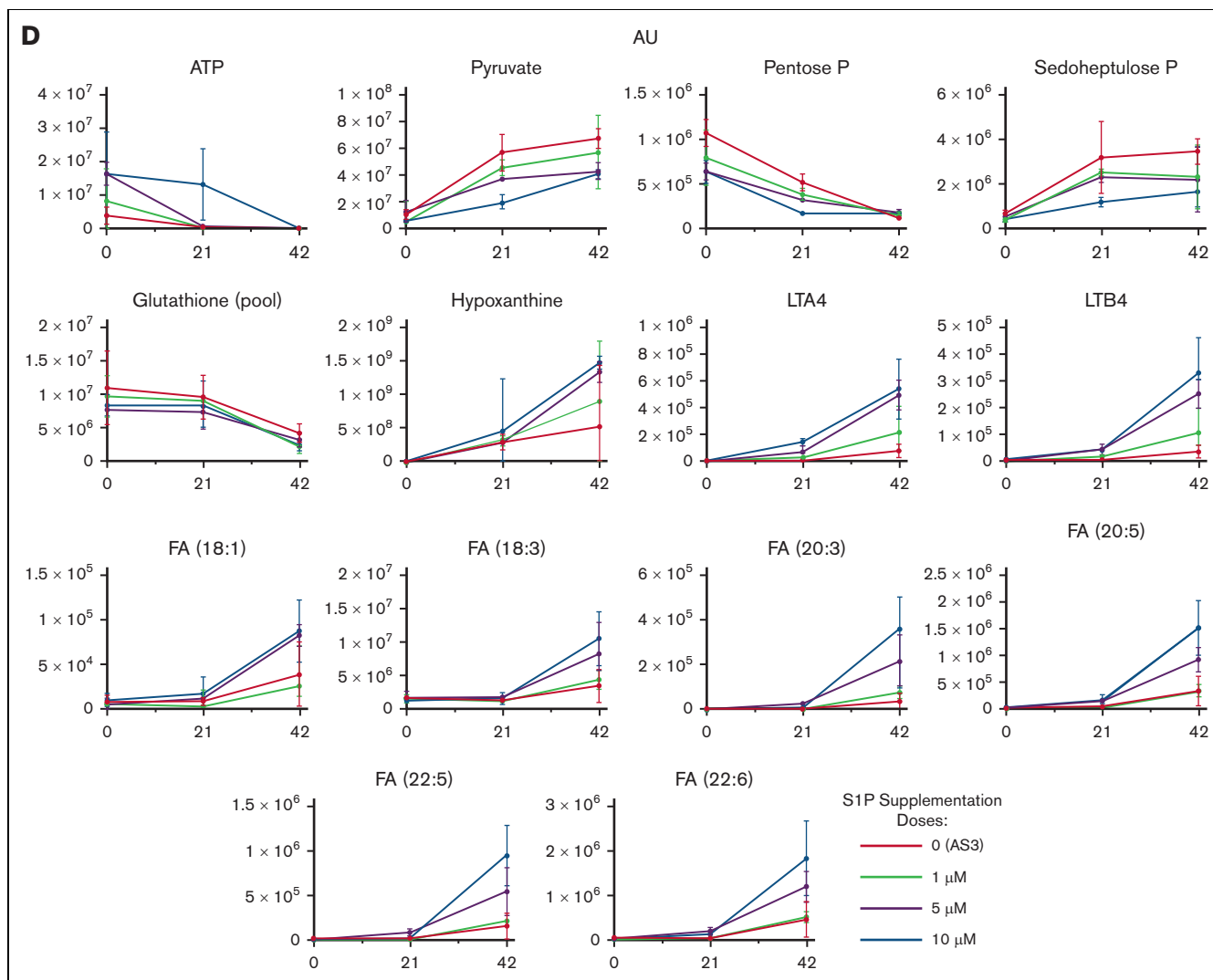
### Storage of human RBCs with S1P promotes glycolysis and generation of ATP, at the expense of steady-state levels of metabolites from the PPP and other antioxidant pathways

Previous studies have shown that quality of RBCs declines with storage, as determined by increased hemolytic propensity<sup>14</sup> and reduction in PTRs.<sup>35,55</sup> Here, S1P decreases were observed as a function of storage duration (Figure 1B). Packed RBCs from female and older donors, all showing higher levels of S1P independent of storage time, have previously been reported to have lower hemolysis,<sup>14</sup> metabolic dysregulation,<sup>8</sup> and promote higher hemoglobin increments upon transfusion.<sup>11,15</sup> Based on these results, we hypothesized that lower levels of RBC S1P are

associated with metabolic impairment of ATP and DPG generation capacity, and that storage quality of human RBCs could be improved by supplementation of S1P to storage additives. To test this hypothesis, human packed RBCs were stored in AS-3, either untreated or supplemented with 1, 5, or 10  $\mu$ M of S1P (Figure 2A). These doses were chosen to recapitulate the ranges in human RBCs exposed to high-altitude hypoxia,<sup>37</sup> while remaining below the critical micelle concentration of S1P in aqueous solution (12  $\mu$ M).<sup>56</sup> Statistical analyses, including time-series ANOVA as a function of S1P supplementation dose, revealed a significant impact of S1P on stored human RBC metabolism and supernatants (heat maps in Figure 2B-C, respectively). Notably, S1P did promote increases in the steady-state levels of ATP and glycolytic metabolites (pyruvate, Figure 2D); however, decreases in the



**Figure 2.** Storage of human RBCs with S1P promotes glycolysis and generation of ATP, at the expense of steady-state levels of metabolites from the PPP and other antioxidant pathways. (A) Human packed RBCs were stored in AS-3, either untreated or supplemented with 1, 5, or 10  $\mu\text{M}$  of S1P. (B-C) Heat maps from metabolomics

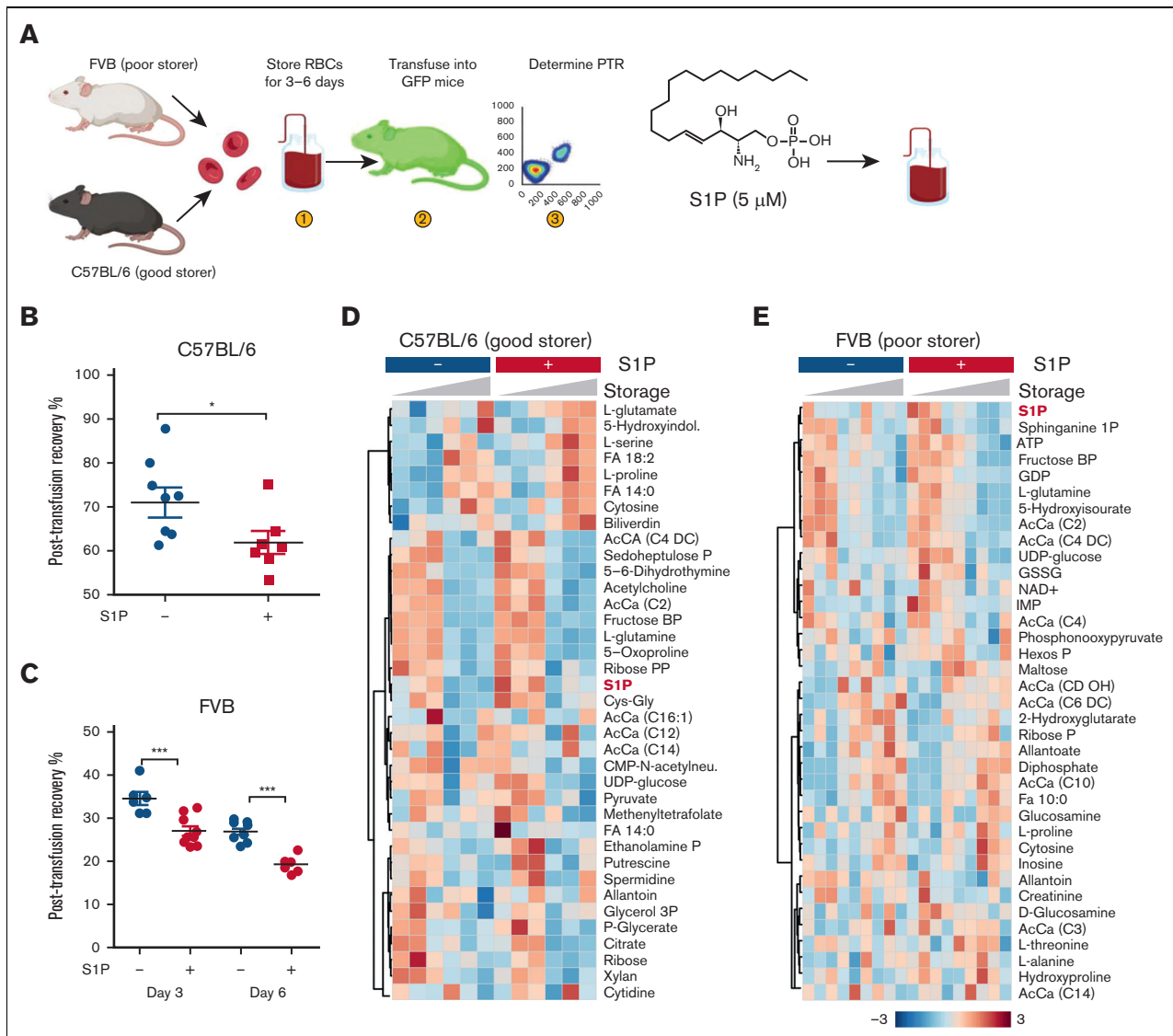


**Figure 2 (continued)** analyses of RBCs and supernatants from this experiment are shown in panels B and C, respectively. (D) Line plots (median  $\pm$  ranges) for representative metabolites in glycolysis (pyruvate), ATP, PPP metabolites (ribose phosphate and pentose phosphate isobars, sedoheptulose phosphate), total glutathione (pools of reduced and oxidized), free fatty acids, and leukotrienes (LTA4 and LTB4). (E) A schematic representation of the results.

steady-state levels of PPP metabolites were observed in a dose-response manner (for example, ribose phosphate and pentose phosphate isomers; sedoheptulose phosphate; [Figure 2D](#)). Consistently, we observed decreases in total glutathione (pools of reduced and oxidized), and increases in free long-chain polyunsaturated and highly unsaturated fatty acids (C18:1, 18:3, 20:3, 20:5, 22:5, and 22:6), markers of oxidant stress-induced fatty acid desaturase activity,<sup>57</sup> as a function of S1P levels ([Figure 2D](#)). Similarly, in S1P-supplemented RBCs we observed an accumulation of oxylipins, including leukotrienes (LTA4 and LTB4) and hydroxyeicosatetraenoic acids, as well as oxidized purines (for example, hypoxanthine), markers of poor PTR in both mice<sup>48</sup> and humans<sup>58</sup> ([Figure 2D](#)). In brief, storage in presence of S1P did boost glycolysis and ATP levels, at the expense of the antioxidant system, that is, the PPP and glutathione pools, and purine and lipid oxidation ([Figure 2E](#)).

## Mouse RBC storage upon supplementation of S1P decreases PTR

To further validate these findings in a tractable animal model, we performed similar storage experiments in mice, in which PTR studies are more easily executed and mechanistic intervention (for example, via genetic ablation of the S1P biosynthetic pathway) are amenable to testing. Cognizant that not all mouse strain RBCs store similarly,<sup>46</sup> we performed follow-up studies on multiple mouse strains. Indeed, some mouse strains have been previously<sup>46,48</sup> labeled as good storers, because of their elevated PTRs (for example, C57BL6/J), whereas other mouse strains are characterized by an exacerbated metabolic storage lesion and poor PTRs (FVB).<sup>46</sup> Therefore, S1P supplementation (5  $\mu$ M) was performed both in good and poor storer strains ([Figure 3A](#)), resulting in lower PTRs in both strain types upon S1P supplementation ([Figure 3B-C](#)). An overview of the



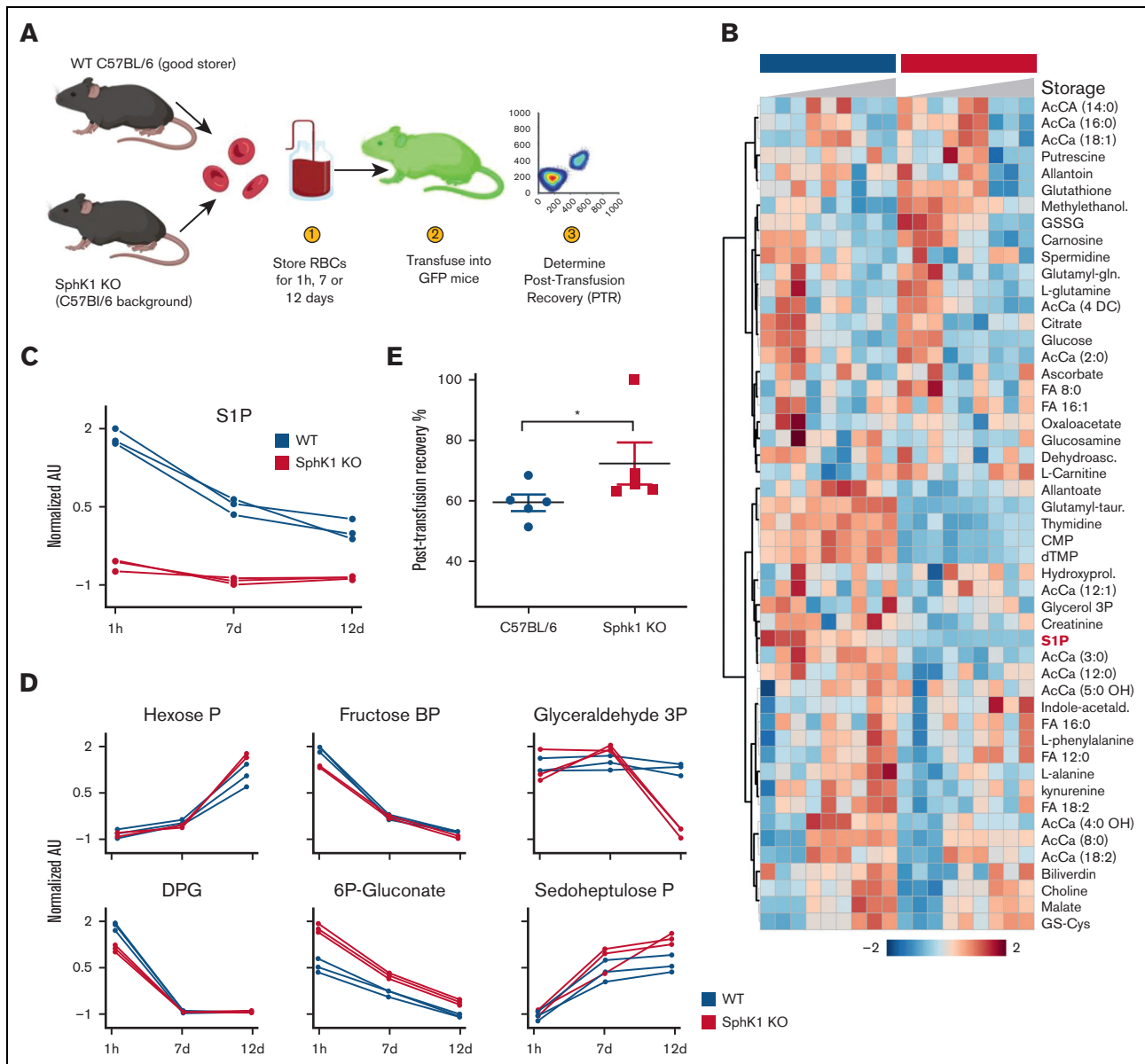
**Figure 3. Mouse RBC storage upon supplementation of S1P decreases post-transfusion recovery.** Post transfusion recovery studies were performed in FVB and C57BL/6 mice upon storage of RBCs for 3 and 6 days in presence or absence of exogenously supplemented S1P (5 μM) (A). Decreases in PTR (median ± ranges) were observed in S1P supplemented mice for 3 and 6 days (B-C). S1P had a measurable effect on stored RBC glycolysis, free and acyl-conjugated carnitines in both good storer C57BL/6 and poor storer FVB mice over storage duration (heat maps in panels D and E, respectively, showing triplicate metabolite measurements at storage days 3 and 6).

murine protocol for measurements of PTR with the combined use of UBC-GFP recipient mice and mCHERRY reporter cells is provided in supplemental Figure 4A-C, along with representative scatter plots from flow cytometry experiments. Poorer storage quality in S1P-supplemented RBCs was associated with a measurable effect on glycolysis, and free and acyl-conjugated carnitines in both good storer C57BL/6 and poor storer FVB mice (heat maps in Figure 3D-E, respectively).

### Genetic ablation of S1P biosynthesis in Sphk1 KO mice is associated with improved storage quality

Given the negative effect of S1P supplementation on human and murine RBC storage quality, we hypothesized that an opposite, beneficial effect would be observed in response to genetic ablation of

the S1P biosynthetic pathway in Sphk1 KO mice. To test this hypothesis, RBCs from wild-type (WT) C57BL/6 and Sphk1 KO mice were stored (Figure 4A), which resulted in a beneficial effect on storage-induced depletion of acylcarnitines and glutathione pools, and mitigation of free fatty acid and purine oxidation product accumulation (allantoate, heat map in Figure 4B). Confirming Sphk1 genetic ablation, significantly lower levels of RBC S1P were observed in these mice (Sphk1 mice had ~30% of S1P levels of WT mice at baseline - Figure 4C). Sphk1 KO stored RBCs were also characterized by lower levels of glycolytic metabolites (glucose-6-phosphate and hexose phosphate isomers, fructose bisphosphate, glyceraldehyde-3-phosphate, and DPG) and higher levels of PPP metabolites (6-phosphogluconate, sedoheptulose phosphate; Figure 4D). Consistent with the beneficial effect of decreased S1P levels, Sphk1



**Figure 4. Improved PTR and antioxidant metabolism in RBCs from Sphk1 KO mice.** (A-C) Storage of mouse RBCs from WT C57BL/6 and Sphk1 KO mice (A) resulted in altered metabolism (heat map in panel B), including significantly lower levels of RBC S1P (C), as expected. (D) Specifically, Sphk1 KO stored RBCs were characterized by lower levels of glycolytic metabolites (hexose phosphate, fructose biphosphate, glyceraldehyde-3-phosphate, and DPG) and higher levels of PPP metabolites (6-phosphogluconate, sedoheptulose phosphate). Sphk1 mice had higher PTR ( $P < .05$ ) than WT good storer mice (median  $\pm$  ranges) (E). In panel E, 1 data point for PTR in Sphk1 mice was set to 100%, as explained in "Materials and methods."

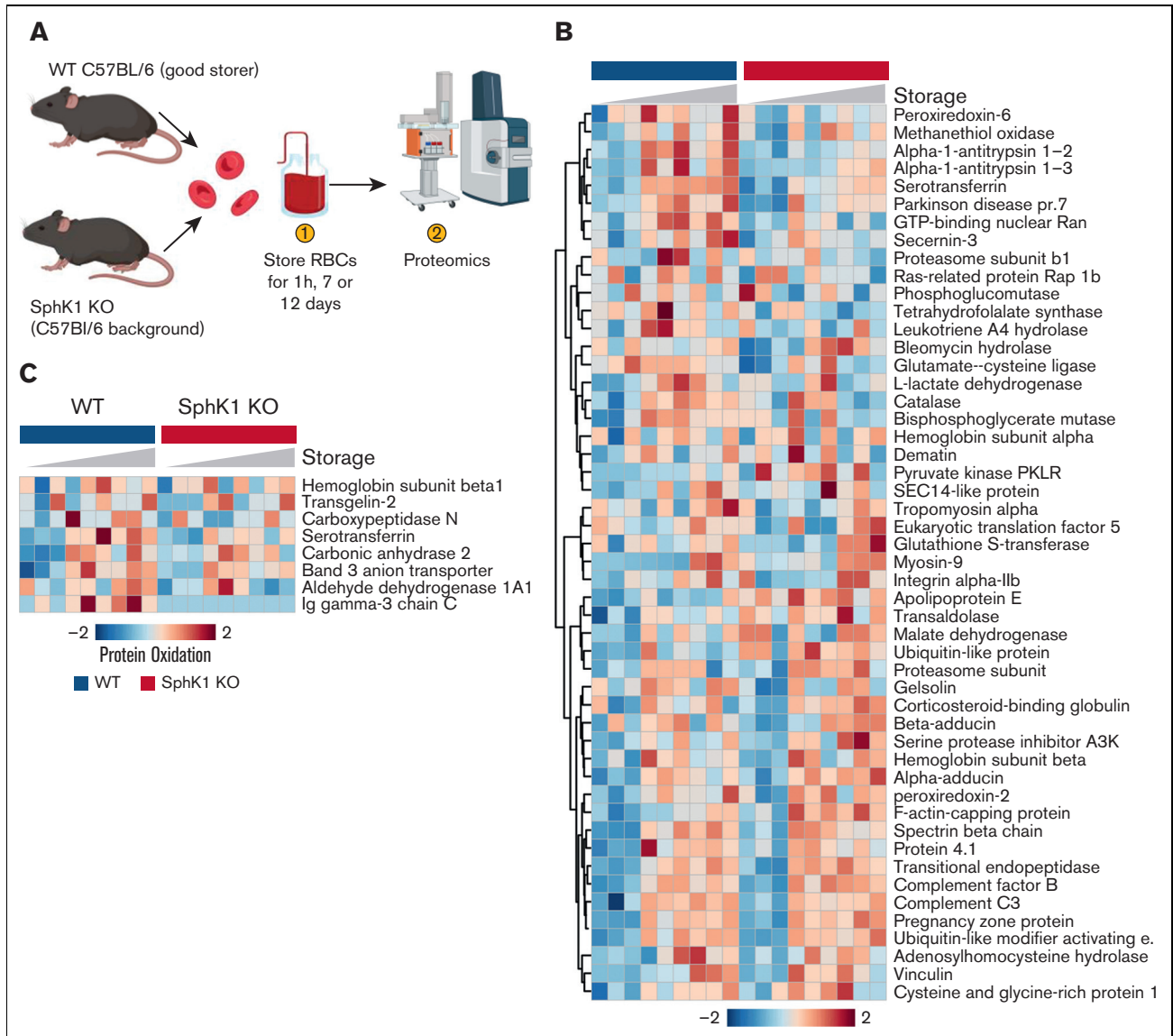
mice (already with a C57BL/6/J good storer background) had higher PTR ( $P < .05$ ) than WT good storer mice (Figure 4E).

Consistent with increased antioxidant capacity and improved metabolic phenotypes and PTRs, Sphk1 mice were characterized by a mitigated storage lesion to the proteome (Figure 5A-B). Specifically, significantly lower levels of oxidized proteins (cysteine and methionine oxidation) were observed in Sphk1 KO mice, including the most abundant cytosolic and membrane proteins, hemoglobin subunit beta 1 and band-3 anion transporter, respectively (Figure 5C).

### Hypoxic storage of Sphk1 KO RBCs normalizes glycolytic fluxes while preserving fluxes through the PPP, thus further boosting PTRs

Hypoxic storage of human<sup>28</sup> and murine (publication forthcoming) RBCs mitigates the metabolic storage lesion and boosts PTRs by promoting glycolysis and mitigating the oxidative storage lesion. Given the apparent effect of Sphk1 on boosting the PPP at the expenses of glycolysis, we hypothesized that altered glucose-oxidation fluxes would be normalized in Sphk1 KO mice by storage in hypoxia (Figure 6A), with consequent benefits on PTRs.





**Figure 5. Impact of Sphk1 KO on the stored RBC proteome.** Stored RBCs from Sphk1 mice (A) have lower alterations to the proteome (heat map in panel B), including protein oxidation (heat map in panel C).

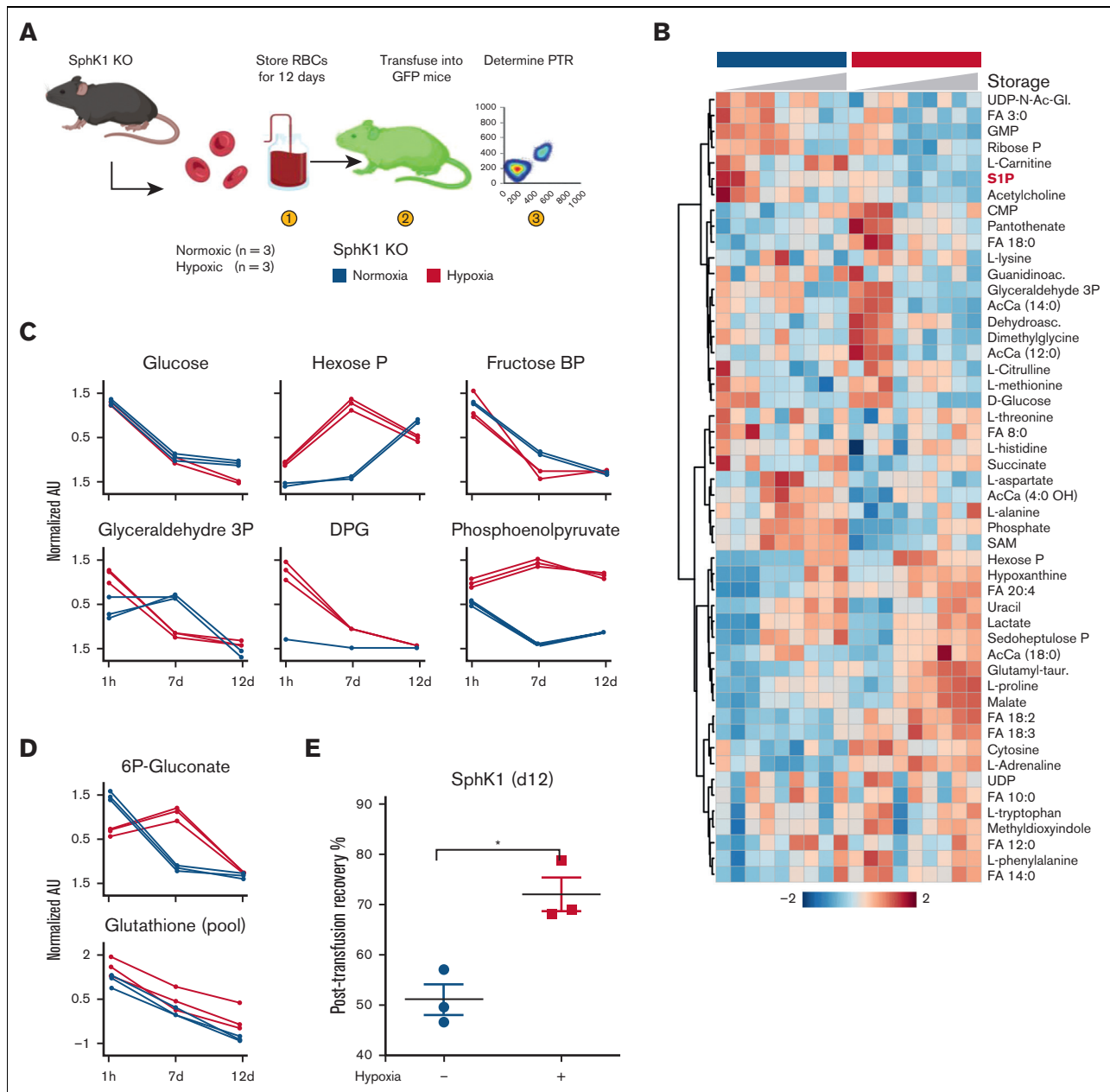
Although metabolic effects of hypoxic storage of WT RBCs have recently been described (publication forthcoming), hypoxic Sphk1 KO were found to have significantly different metabolism (Figure 6B). Specifically, compared with their normoxic counterparts, hypoxic Sphk1 KO RBCs had higher glucose consumption, as well as increased levels of multiple glycolytic metabolites (hexose phosphate, DPG, phosphoenolpyruvate; Figure 6C). These changes were not accompanied by decreases in the levels of PPP metabolites (6-phosphogluconate) and total glutathione pools, which were higher in the hypoxic Sphk1 KO mouse RBCs (Figure 6D). These metabolic effects were accompanied by significant improvements in PTRs (Figure 6E).

To further confirm the metabolic mechanism driving these observations, we performed tracing experiments with 1,2,3-<sup>13</sup>C<sub>3</sub>-glucose (Figure 7A). This approach affords discrimination of metabolic

fluxes through glycolysis and the PPP (Figure 7B) via determination of the ratio of +3 and +2 lactate isotopologues (Figure 7C), as previously described.<sup>27,32,33</sup> Results clearly indicate that, compared with WT normoxic RBCs, hypoxic WT RBCs had significantly elevated lactate +3 and glycolysis/PPP ratios (Figure 7D). Sphk1 KO mice had slower fluxes through glycolysis compared with WT normoxic and hypoxic RBCs. However, hypoxic Sphk1 KO RBCs had comparable fluxes through glycolysis to WT normoxic RBCs (Figure 7D), despite lower overall storage-induced accumulation of markers of oxidant stress (Figure 6).

## Discussion

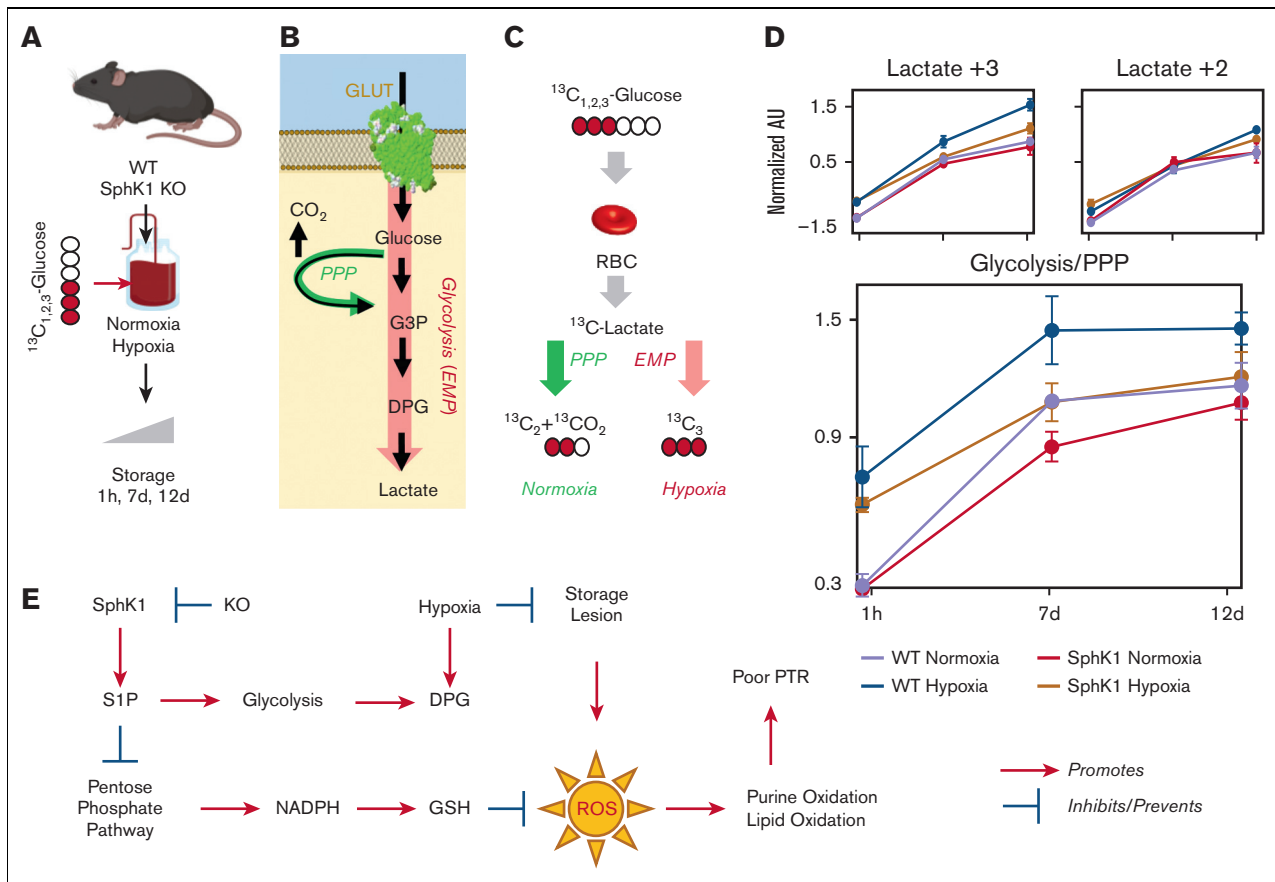
In this study, we delved into the role of S1P in RBC storage quality. The underlying hypothesis was that storage depletes ATP and DPG pools, and that S1P can boost ATP and DPG synthesis by



**Figure 6. Impact of hypoxic storage and Sphk1 KO on the stored RBC metabolome.** Storage of Sphk1 mouse RBCs under hypoxic conditions (A) further improves the metabolic benefits (heat map in B) of either treatment alone by boosting glycolysis (C), glutathione pools without depressing the PPP (D) and PTR (median  $\pm$  ranges) (E).

promoting metabolic fluxes through glycolysis.<sup>37</sup> Because the literature was controversial on the impact of storage on RBC S1P levels, either reporting declines,<sup>39</sup> no change,<sup>40</sup> or increases with some additives,<sup>41</sup> we performed S1P measurements on, to our knowledge, the largest longitudinal sample set of stored RBC samples available to date (~2000 samples from 643 donors at 3 time points) to confirm that S1P levels do decrease with storage. Notably, we also observed higher levels of S1P, independent of storage duration, in better-storing units (donors with lower hemolytic propensity<sup>14</sup>), including units from female and older donors. To test the hypothesis that S1P improves RBC storage quality, we either supplemented human and murine RBCs with S1P before

storage, or relied on mice lacking the capacity to synthesize S1P (Sphk1 KO). As a result, PTR, proteomics, and steady state and tracing metabolomics experiments all suggest that S1P does indeed boost glycolysis and ATP and DPG generation in stored human and murine RBCs; however, it does so at the expenses of metabolic fluxes through the PPP. This metabolic rewiring comes at the cost of a decreased antioxidant capacity and increased storage-induced oxidation of proteins, lipids, and purines. Indeed, S1P supplementation was associated with increases in the levels of (i) purine oxidation products (hypoxanthine), the result of oxidative stress-activated adenosine monophosphate deaminase 3<sup>58</sup>; (ii) free polyunsaturated fatty acids, the result of oxidant



**Figure 7. Sphk1 and hypoxia impact metabolic fluxes through glycolysis and the pentose phosphate pathway in stored murine RBCs.** Tracing experiments with 1,2,3-<sup>13</sup>C<sub>3</sub>-glucose in stored RBCs from WT and Sphk1 mice (A) afford discrimination of metabolic fluxes through glycolysis and the PPP (B) via determination of the ratio of +3 and +2 lactate isotopologues (C). Increases in glycolysis were observed in hypoxia in WT mice but not in Sphk1 mice. (D) However, glycolysis/PPP ratios were basally lower in Sphk1 mice than in controls and were increased by storage in hypoxia in both groups (despite Sphk1 preserving higher PPP activation compared with WT) (median ± ranges). (E) A summary overview of the main findings of this study is provided.

stress-induced fatty acid desaturase activation<sup>57</sup>; (iii) oxylipins, the result of lipid peroxidation events triggered by dysregulated iron metabolism<sup>48</sup>; and (iv) thiol groups, histidine and asparagine oxidation in hemoglobin (at cysteine beta 93)<sup>59,60</sup> and band 3,<sup>26</sup> all critical regulators of RBC function and structure. Of note, all of these aforementioned lesions have previously been associated with poorer PTRs,<sup>32,48,57-60</sup> which is what we observed in S1P-supplemented murine units. Genetic manipulation of Sphk1,<sup>45</sup> the rate-limiting step in S1P biosynthesis in mature RBCs, counteracts these phenomena, whereas hypoxia boosts glycolysis while normalizing fluxes through glycolysis in Sphk1 KO mice and preventing oxidative metabolic lesions (findings summarized in the schematic in Figure 7E).

Notably, our results suggest that S1P supplementation phenocopies G6PD deficiency, in which a boost in energy metabolism is accompanied by suppressed fluxes through the PPP; as a result G6PD-deficient RBCs are characterized by an increase in oxidant stress to metabolites, proteins, and lipids,<sup>7,8</sup> and a negative impact on PTRs.<sup>10</sup> Because G6PD activity (and thus PPP fluxes) have previously been reported to decline with age,<sup>12</sup> it is interesting to note that, in this study, S1P levels were observed to be positively

correlated with donor age. However, it must be noted that healthy, frequent blood donors were enrolled in the REDS study, which biases the population toward subjects with lower propensity to hemolyze following oxidant insults, as reported.<sup>61</sup> In addition, because higher G6PD expression and activity levels are associated with longer lifespan,<sup>13</sup> a survival bias may exist in older donors who are healthy enough to donate blood frequently. In female blood donors, the negative effect of S1P on PPP activation may be instead counteracted by increased chromosome X-linked G6PD protein dosage, whereby RBC protein levels and activity are higher compared with males despite chromosome-X inactivation.<sup>62</sup> In addition, it is worth noting that prior studies on the blood donor exposome have shown how over-the-counter drugs that are not grounds for blood donor deferral (for example, proton pump inhibitors) can boost RBC S1P levels.<sup>63</sup> Similarly, it is worth noting that although no genetic polymorphism in Sphk1 was observed to be associated with RBC hemolytic propensity in prior large GWAS studies (for example, REDS III),<sup>9</sup> variation in the MFSD2b gene<sup>64</sup> (an S1P transporter) was significantly associated with osmotic fragility of the stored RBC.<sup>9</sup> Indeed, erythrocytes efficiently utilize exogenous sphingosines for S1P synthesis and export via Mfsd2b.<sup>44</sup> The combination of sphingolipid uptake, here validated with deuterium-

labeled S1P dose-response incubation experiments, and RBC-specific<sup>37</sup> Sphk1 activity regulates not just intraerythrocytic levels of S1P, with RBCs de facto acting as reservoirs that modulate circulating concentrations of S1P.<sup>43</sup>

Of note, hypoxic storage of Sphk1 RBC boosted energy metabolism while preventing the accumulation of oxidative stress markers. Hypoxic storage improved PTRs of both Sphk1 and WT mice, observation in the latter confirming recent findings in mice.<sup>47</sup> This observation is interesting in that we had hypothesized a role for S1P in stabilizing deoxyhemoglobin and promoting glycolysis. However, in the Sphk1 KO mouse the stabilizing effect of S1P on deoxyhemoglobin is limited, which indeed corresponded to slower fluxes through glycolysis compared with WT mice upon exposure to hypoxia. Alkalinization of the intracellular milieu under hypoxic storage conditions had been suggested to boost the activity of phosphofructokinase, biphosphoglycerate mutase, and G6PD, the rate-limiting enzymes of glycolysis, the Rapoport-Luebering shunt, and the PPP, respectively.<sup>65</sup> Similar beneficial effects on all 3 pathways are indeed observed when RBCs are stored with alkaline additives.<sup>66,67</sup> As such, this study provides further evidence that the pH effect and the oxygen-dependent metabolic modulation model proposed by multiple groups over the years<sup>31-33,68</sup> are mutually relevant in the economy of RBC energy/redox glucose-oxidation fluxes, and that S1P does play a non-secondary role in this system. Notably, storage-induced fragmentation of the N-terminus cytosolic domain of band 3,<sup>32,33,69</sup> because of radical attack or caspase activity,<sup>70,71</sup> ultimately impairs the RBC capacity to respond to oxidant stress by activating the PPP as a function of high SO<sub>2</sub>, making S1P-induced promotion of deoxyhemoglobin interaction with the very same N-terminus of band 3<sup>37</sup> less mechanistically relevant. In light of the decreased stress to band 3 under hypoxic storage conditions,<sup>26</sup> it is interesting to hypothesize that S1P supplementation in hypoxically stored erythrocytes may still represent a viable storage strategy to further enhance the metabolic and posttransfusion benefits of these products.<sup>28</sup> Similar considerations can be made for alkaline additives.<sup>66,67</sup>

It should be noted that this study focuses on the stored RBCs and their posttransfusion performances in heterologous, healthy murine recipients. As such, it does not take into account nonhealthy, hypoxic transfusion recipients. Because S1P is depleted in patients that are hypoxic with trauma/hemorrhagic shock<sup>72</sup> or with sepsis,<sup>73</sup> S1P-containing additives may boost oxygen kinetics in this recipient population and phenocopy the beneficial effects of S1P agonists like FTY720.<sup>74</sup> In contrast, in certain categories of recipients, like patients with sickle cell disease, stabilization of deoxy-sickle hemoglobin is potentially deleterious because it would trigger polymerization and sickling.<sup>36</sup> All these considerations will be

critical if further investigations on the use of S1P as a supplement to blood products are pursued.

## Acknowledgments

Research reported in this publication was supported by funds from R01HL146442, R01HL149714, and R01HL148151 (A.D. and J.C.Z.) and R21HL150032 (A.D.) by the National Heart, Lung, and Blood Institute. The REDS-IV-P CTLS project was sponsored by the National Heart, Lung, and Blood Institute contract 75N2019D00033. Samples were collected as part of the NHLBI Recipient Epidemiology and Donor Evaluation Study-III (REDS-III) RBC Omics project, which was supported by National Heart, Lung, and Blood Institute contracts NHLBI HHSN2682011-000011, -000021, -000031, -000041, -000051, -000061, -000071, -000081, and -000091.

## Authorship

Contribution: A.D., Y.X., and J.C.Z. designed the study; A.H. performed mouse work and performed mouse PTR studies, and together with J.C.Z. interpreted the results; A.H. and J.C.Z. generated the ROSA26-LCB-mCherry mouse; F.G., T.N., M.D., A.K., and A.D. performed omics analyses; M.P.B. designed and supervised the REDS-III RBC Omics study; K.B. performed bioinformatics analyses; A.D. performed data analysis and prepared figures and tables; A.D. wrote the first draft of the manuscript; and all authors contributed to manuscript revisions, editing, and finalization of the manuscript.

Conflict-of-interest disclosure: The authors declare that A.D. and T.N. are founders of Omix Technologies Inc. A.D. is also a consultant for Rubius Inc and Forma Inc. A.D. is an advisory board member of Macopharma and Hemanext Inc, a company that is developing a product for hypoxic storage of human RBCs. J.C.Z. is a consultant for Rubius Inc, and is a cofounder and the chief scientific officer for Svalinn Therapeutics, a company whose focus is unrelated to the current work. A.D., Y.X., and T.N. hold a patent on S1P supplementation on stored RBC units. The remaining authors declare no competing financial interests.

ORCID profiles: T.N., 0000-0001-8566-7119; A.K., 0000-0002-8787-8144; Y.X., 0000-0002-8059-6179; P.J.N., 0000-0003-0526-2088; A.D., 0000-0002-2258-6490.

Correspondence: Angelo D'Alessandro, Department of Biochemistry and Molecular Genetics, University of Colorado Anschutz Medical Campus, 12801 East 17th Ave, Aurora, CO 80045; email: [angelo.dalessandro@cuanschutz.edu](mailto:angelo.dalessandro@cuanschutz.edu).

## References

1. Roussel C, Morel A, Dussiot M, et al. Rapid clearance of storage-induced microerythrocytes alters transfusion recovery. *Blood*. 2021;137(17):2285-2298.
2. D'Alessandro A, D'Amici GM, Vaglio S, Zolla L. Time-course investigation of SAGM-stored leukocyte-filtered red blood cell concentrates: from metabolism to proteomics. *Haematologica*. 2012;97(1):107-115.
3. Yoshida T, Prudent M, D'Alessandro A. Red blood cell storage lesion: causes and potential clinical consequences. *Blood Transfus*. 2019;17(1):27-52.
4. Kanias T, Acker JP. Biopreservation of red blood cells—the struggle with hemoglobin oxidation. *FEBS J*. 2010;277(2):343-356.

5. D'Alessandro A, Hansen KC, Eisenmesser EZ, Zimring JC. Protect, repair, destroy or sacrifice: a role of oxidative stress biology in inter-donor variability of blood storage? *Blood Transfus.* 2019;17(4):281-288.
6. Luzzatto L, Ally M, Notaro R. Glucose-6-phosphate dehydrogenase deficiency. *Blood.* 2020;136(11):1225-1240.
7. Tzounakas VL, Kriebardis AG, Georgatzakou HT, et al. Glucose 6-phosphate dehydrogenase deficient subjects may be better "storsers" than donors of red blood cells. *Free Radic Biol Med.* 2016;96:152-165.
8. D'Alessandro A, Fu X, Kanas T, et al. Donor sex, age and ethnicity impact stored red blood cell antioxidant metabolism through mechanisms in part explained by glucose 6-phosphate dehydrogenase levels and activity. *Haematologica.* 2021;106(5):1290-1302.
9. Page GP, Kanas T, Guo YJ, et al. Multiple-ancestry genome-wide association study identifies 27 loci associated with measures of hemolysis following blood storage. *J Clin Invest.* 2021;131(13):e146077.
10. Francis RO, D'Alessandro A, Eisenberger A, et al. Donor glucose-6-phosphate dehydrogenase deficiency decreases blood quality for transfusion. *J Clin Invest.* 2020;130(5):2270-2285.
11. Roubinian NH, Reese SE, Qiao H, et al. Donor genetic and nongenetic factors affecting red blood cell transfusion effectiveness. *JCI Insight.* 2022;7(1):e152598.
12. Rodgers GP, Lichtman HC, Sheff MF. Red blood cell glucose-6-phosphate dehydrogenase activity in aged humans. *J Am Geriatr Soc.* 1983;31(1):8-11.
13. Nóbrega-Pereira S, Fernandez-Marcos PJ, Brioché T, et al. G6PD protects from oxidative damage and improves healthspan in mice. *Nat Commun.* 2016;7(1):10894.
14. Kanas T, Lanteri MC, Page GP, et al. Ethnicity, sex, and age are determinants of red blood cell storage and stress hemolysis: results of the REDS-III RBC-Omics study. *Blood Adv.* 2017;1(15):1132-1141.
15. Roubinian NH, Plimier C, Woo JP, et al. Effect of donor, component, and recipient characteristics on hemoglobin increments following red blood cell transfusion. *Blood.* 2019;134(13):1003-1013.
16. D'Alessandro A, Zimring JC, Busch M. Chronological storage age and metabolic age of stored red blood cells: are they the same? *Transfusion.* 2019;59(5):1620-1623.
17. Koch CG, Duncan AI, Figueroa P, et al. Real age: red blood cell aging during storage. *Ann Thorac Surg.* 2019;107(3):973-980.
18. Hess JR. An update on solutions for red cell storage. *Vox Sang.* 2006;91(1):13-19.
19. Bogdanova A, Makhro A, Wang J, Lipp P, Kaestner L. Calcium in red blood cells—a perilous balance. *Int J Mol Sci.* 2013;14(5):9848-9872.
20. Nemkov T, Qadri SM, Sheffield WP, D'Alessandro A. Decoding the metabolic landscape of pathophysiological stress-induced cell death in anucleate red blood cells. *Blood Transfus.* 2020;18(2):130-142.
21. Rinalducci S, Longo V, Ceci LR, Zolla L. Targeted quantitative phosphoproteomic analysis of erythrocyte membranes during blood bank storage. *J Mass Spectrom.* 2015;50(2):326-335.
22. Donovan K, Meli A, Cendali F, et al. Stored blood has compromised oxygen unloading kinetics that can be normalized with rejuvenation and predicted from corpuscular side-scatter. *Haematologica.* 2021;107(1):298-302.
23. van Wijk R, van Solinge WW. The energy-less red blood cell is lost: erythrocyte enzyme abnormalities of glycolysis. *Blood.* 2005;106(13):4034-4042.
24. Ahmed MH, Ghatge MS, Safo MK. Hemoglobin: structure, function and allostery. *Subcell Biochem.* 2020;94:345-382.
25. Yoshida T, McMahon E, Croxon H, et al. The oxygen saturation of red blood cell concentrates: the basis for a novel index of red cell oxidative stress. *Transfusion.* 2022;62(1):183-193.
26. Reisz JA, Nemkov T, Dzieciatkowska M, et al. Methylation of protein aspartates and deamidated asparagines as a function of blood bank storage and oxidative stress in human red blood cells. *Transfusion.* 2018;58(12):2978-2991.
27. Reisz JA, Wither MJ, Dzieciatkowska M, et al. Oxidative modifications of glyceraldehyde 3-phosphate dehydrogenase regulate metabolic reprogramming of stored red blood cells. *Blood.* 2016;128(12):e32-42.
28. D'Alessandro A, Yoshida T, Nestheide S, et al. Hypoxic storage of red blood cells improves metabolism and post-transfusion recovery. *Transfusion.* 2020;60(4):786-798.
29. Williams AT, Jani VP, Nemkov T, et al. Transfusion of anaerobically or conventionally stored blood after hemorrhagic shock. *Shock.* 2020;53(3):352-362.
30. Dumont LJ, D'Alessandro A, Szczepiorkowski ZM, Yoshida T. CO<sub>2</sub> -dependent metabolic modulation in red blood cells stored under anaerobic conditions. *Transfusion.* 2016;56(2):392-403.
31. Lewis IA, Campanella ME, Markley JL, Low PS. Role of band 3 in regulating metabolic flux of red blood cells. *Proc Natl Acad Sci U S A.* 2009;106(44):18515-18520.
32. Issaian A, Hay A, Dzieciatkowska M, et al. The interactome of the N-terminus of band 3 regulates red blood cell metabolism and storage quality. *Haematologica.* 2021;106(11):2971-2985.
33. Rogers SC, Ge X, Brummet M, et al. Quantifying dynamic range in red blood cell energetics: evidence of progressive energy failure during storage. *Transfusion.* 2021;61(5):1586-1599.
34. Rabcuka J, Blonski S, Meli A, et al. Metabolic reprogramming under hypoxic storage preserves faster oxygen unloading from stored red blood cells. *Blood Adv.* 2022;6(18):5415-5428.

35. de Bruin S, Peters A-L, Wijnberge M, et al. Storage of red blood cells in alkaline PAGGGM improves metabolism but has no effect on recovery after transfusion. *Blood Adv.* 2022;6(13):3899-3910.
36. Sun K, D'Alessandro A, Ahmed MH, et al. Structural and functional insight of sphingosine 1-phosphate-mediated pathogenic metabolic reprogramming in sickle cell disease. *Sci Rep.* 2017;7(1):15281.
37. Sun K, Zhang Y, D'Alessandro A, et al. Sphingosine-1-phosphate promotes erythrocyte glycolysis and oxygen release for adaptation to high-altitude hypoxia. *Nat Commun.* 2016;7:12086.
38. Xie T, Chen C, Peng Z, et al. Erythrocyte metabolic reprogramming by sphingosine 1-phosphate in chronic kidney disease and therapies. *Circ Res.* 2020;127(3):360-375.
39. Selim S, Sunkara M, Salous AK, et al. Plasma levels of sphingosine 1-phosphate are strongly correlated with haematocrit, but variably restored by red blood cell transfusions. *Clin Sci (Lond).* 2011;121(12):565-572.
40. Poppe A, Moritz E, Geffken M, et al. Analyses of sphingosine-1-phosphate in the context of transfusion: how much is in stored blood products and in patient blood? *Transfusion.* 2019;59(10):3071-3076.
41. D'Alessandro A, Culp-Hill R, Reisz JA, et al. Heterogeneity of blood processing and storage additives in different centers impacts stored red blood cell metabolism as much as storage time: lessons from REDS-III-Omics. *Transfusion.* 2019;59(1):89-100.
42. Endres-Dighe SM, Guo Y, Kanas T, et al. Blood, sweat, and tears: Red Blood Cell-Omics study objectives, design, and recruitment activities. *Transfusion.* 2019;59(1):46-56.
43. Bode C, Sensken SC, Peest U, et al. Erythrocytes serve as a reservoir for cellular and extracellular sphingosine 1-phosphate. *J Cell Biochem.* 2010;109(6):1232-1243.
44. Nguyen TQ, Vu TM, Tukijan F, et al. Erythrocytes efficiently utilize exogenous sphingosines for S1P synthesis and export via Mfsd2b. *J Biol Chem.* 2021;296:100201.
45. Allende ML, Sasaki T, Kawai H, et al. Mice deficient in sphingosine kinase 1 are rendered lymphopenic by FTY720. *J Biol Chem.* 2004;279(50):52487-52492.
46. Zimring JC, Smith N, Stowell SR, et al. Strain-specific red blood cell storage, metabolism, and eicosanoid generation in a mouse model. *Transfusion.* 2014;54(1):137-148.
47. Hay A, Dziewulska KH, Gamboni F, et al. Hypoxic storage of murine red blood cells improves energy metabolism and post-transfusion recoveries. *Blood Transfus.* Published online 21 October 2022.
48. Howie HL, Hay AM, de Wolski K, et al. Differences in Steap3 expression are a mechanism of genetic variation of RBC storage and oxidative damage in mice. *Blood Adv.* 2019;3(15):2272-2285.
49. Nemkov T, Hansen KC, D'Alessandro A. A three-minute method for high-throughput quantitative metabolomics and quantitative tracing experiments of central carbon and nitrogen pathways. *Rapid Commun Mass Spectrom.* 2017;31(8):663-673.
50. Nemkov T, Yoshida T, Nikulina M, D'Alessandro A. High-throughput metabolomics platform for the rapid data-driven development of novel additive solutions for blood storage. *Front Physiol.* 2022;13:833242.
51. Nemkov T, Reisz JA, Gehrke S, Hansen KC, D'Alessandro A. High-throughput metabolomics: isocratic and gradient mass spectrometry-based methods. *Methods Mol Biol.* 2019;1978:13-26.
52. Reisz JA, Zheng C, D'Alessandro A, Nemkov T. Untargeted and semi-targeted lipid analysis of biological samples using mass spectrometry-based metabolomics. *Methods Mol Biol.* 2019;1978:121-135.
53. Clasquin MF, Melamud E, Rabinowitz JD. LC-MS data processing with MAVEN: a metabolomic analysis and visualization engine. *Curr Protoc Bioinformatics.* 2012; Chapter 14:Unit14.11.
54. Pang Z, Chong J, Zhou G, et al. MetaboAnalyst 5.0: narrowing the gap between raw spectra and functional insights. *Nucleic Acids Res.* 2021;49(W1):W388-w396.
55. Dumont LJ, AuBuchon JP. Evaluation of proposed FDA criteria for the evaluation of radiolabeled red cell recovery trials. *Transfusion.* 2008;48(6):1053-1060.
56. Garcia-Pacios M, Collado MI, Busto JV, et al. Sphingosine-1-phosphate as an amphipathic metabolite: its properties in aqueous and membrane environments. *Biophys J.* 2009;97(5):1398-1407.
57. Thomas T, Cendali F, Fu X, et al. Fatty acid desaturase activity in mature red blood cells and implications for blood storage quality. *Transfusion.* 2021;61(6):1867-1883.
58. Nemkov T, Sun K, Reisz JA, et al. Hypoxia modulates the purine salvage pathway and decreases red blood cell and supernatant levels of hypoxanthine during refrigerated storage. *Haematologica.* 2018;103(2):361-372.
59. Wither M, Dzieciatkowska M, Nemkov T, Strop P, D'Alessandro A, Hansen KC. Hemoglobin oxidation at functional amino acid residues during routine storage of red blood cells. *Transfusion.* 2016;56(2):421-426.
60. Harper VM, Oh JY, Stapley R, et al. Peroxiredoxin-2 recycling is inhibited during erythrocyte storage. *Antioxid Redox Signal.* 2015;22(4):294-307.
61. Kanas T, Stone M, Page GP, et al. Frequent blood donations alter susceptibility of red blood cells to storage- and stress-induced hemolysis. *Transfusion.* 2019;59(1):67-78.
62. Nantakomol D, Paul R, Palasuwan A, Day NP, White NJ, Imwong M. Evaluation of the phenotypic test and genetic analysis in the detection of glucose-6-phosphate dehydrogenase deficiency. *Malar J.* 2013;12:289.

63. Nemkov T, Stefanoni D, Bordbar A, et al. Blood donor exposome and impact of common drugs on red blood cell metabolism. *JCI Insight*. 2020;6(3):e146175.
64. Vu TM, Ishizu A-N, Foo JC, et al. Mfsd2b is essential for the sphingosine-1-phosphate export in erythrocytes and platelets. *Nature*. 2017;550(7677):524-528.
65. Nemkov T, Hansen KC, Dumont LJ, D'Alessandro A. Metabolomics in transfusion medicine. *Transfusion*. 2016;56(4):980-993.
66. Hess JR, Rugg N, Knapp AD, Gormas JF, Silberstein EB, Greenwalt TJ. Successful storage of RBCs for 10 weeks in a new additive solution. *Transfusion*. 2000;40(8):1012-1016.
67. D'Alessandro A, Reisz JA, Culp-Hill R, Korsten H, van Bruggen R, de Korte D. Metabolic effect of alkaline additives and guanosine/gluconate in storage solutions for red blood cells. *Transfusion*. 2018;58(8):1992-2002.
68. Messana I, Ferroni L, Misiti F, et al. Blood bank conditions and RBCs: the progressive loss of metabolic modulation. *Transfusion*. 2000;40(3):353-360.
69. Lange PF, Huesgen PF, Nguyen K, Overall CM. Annotating N termini for the human proteome project: N termini and N $\alpha$ -acetylation status differentiate stable cleaved protein species from degradation remnants in the human erythrocyte proteome. *J Proteome Res*. 2014;13(4):2028-2044.
70. Rinalducci S, Ferru E, Blasi B, Turrini F, Zolla L. Oxidative stress and caspase-mediated fragmentation of cytoplasmic domain of erythrocyte band 3 during blood storage. *Blood Transfus*. 2012;10(suppl 2):s55-62.
71. Kriebardis AG, Antonelou MH, Stamoulis KE, Economou-Petersen E, Margaritis LH, Papassideri IS. Storage-dependent remodeling of the red blood cell membrane is associated with increased immunoglobulin G binding, lipid raft rearrangement, and caspase activation. *Transfusion*. 2007;47(7):1212-1220.
72. D'Alessandro A, Nemkov T, Moore HB, et al. Metabolomics of trauma-associated death: shared and fluid-specific features of human plasma vs lymph. *Blood Transfus*. 2016;14(2):185-194.
73. Winkler MS, Märtz KB, Nierhaus A, et al. Loss of sphingosine 1-phosphate (S1P) in septic shock is predominantly caused by decreased levels of high-density lipoproteins (HDL). *J Intensive Care*. 2019;7(7):23.
74. Bonitz JA, Son JY, Chandler B, et al. A sphingosine-1 phosphate agonist (FTY720) limits trauma/hemorrhagic shock-induced multiple organ dysfunction syndrome. *Shock*. 2014;42(5):448-455.